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(54) Title: ISOLATED NONAPEPTIDE DERIVED FROM MAGE-3 GENE AND PRESENTED BY HLA-A1, AND USES THEREOF (57) Abstract The invention involves a nonapeptide derived from the tumor rejection antigen precursor encoded by gene MAGE-3. This nonapeptide is presented by HLA molecules HLA-A1. The resulting complexes are identified by cytolytic T cells. Such recognition may be used in diagnostics, or therapeutically.		

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**ISOLATED N NAPEPTIDE DERIVED FR M MAGE-3 GENE
AND PRESENTED BY HLA-A1, AND USES THEREOF**

5 This application is a continuation-in-part of copending application Serial Number 07/938,334, filed August 31, 1992, and Serial Number 08/037,230 filed March 26, 1993.

FIELD OF THE INVENTION

10 This invention relates to immunogenetics and to peptide chemistry. More particularly, it relates to a nonapeptide useful in various ways, including as an immunogen and as a target for the HLA-A1 molecule. More particularly, it relates to a so-called "tumor rejection antigen", derived from the tumor rejection antigen precursor encoded by gene MAGE-3, and presented by human leukocyte antigens of HLA-A1.

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BACKGROUND AND PRIOR ART

The study of the recognition or lack of recognition of cancer cells by a host organism has proceeded in many different directions. Understanding of the field presumes some understanding of both basic immunology and oncology.

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Early research on mouse tumors revealed that these displayed molecules which led to rejection of tumor cells when transplanted into syngeneic animals. These molecules are "recognized" by T-cells in the recipient animal, and provoke a cytolytic T-cell response with lysis of the transplanted cells. This evidence was first obtained with tumors induced in vitro by chemical carcinogens, such as methylcholanthrene. The antigens expressed by the tumors and which elicited the T-cell response were found to be different for each tumor. See Prehn, et al., J. Natl. Canc. Inst. 18: 769-778 (1957); Klein et al., Cancer Res. 20: 1561-1572 (1960); Gross, Cancer Res. 3: 326-333 (1943), Basombrio, Cancer Res. 30: 2458-2462 (1970) for general teachings on inducing tumors with chemical carcinogens and differences in cell surface antigens. This class of antigens has come to be known as "tumor specific transplantable antigens" or "TSTAs". Following the observation of the presentation of such antigens when induced

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by chemical carcinogens, similar results were obtained when tumors were induced in vitro via ultraviolet radiation. See Kripke, J. Natl. Canc. Inst. 53: 333-1336 (1974).

While T-cell mediated immune responses were observed for the types of tumor described supra, spontaneous tumors were thought to be generally non-immunogenic. These were therefore believed not to present antigens which provoked a response to the tumor in the tumor carrying subject. See Hewitt, et al., Brit. J. Cancer 33: 241-259 (1976).

The family of tum⁻ antigen presenting cell lines are immunogenic variants obtained by mutagenesis of mouse tumor cells or cell lines, as described by Boon et al., J. Exp. Med. 152: 1184-1193 (1980), the disclosure of which is incorporated by reference. To elaborate, tum⁻ antigens are obtained by mutating tumor cells which do not generate an immune response in syngeneic mice and will form tumors (i.e., "tum⁺" cells). When these tum⁺ cells are mutagenized, they are rejected by syngeneic mice, and fail to form tumors (thus "tum⁻"). See Boon et al., Proc. Natl. Acad. Sci. USA 74: 272 (1977), the disclosure of which is incorporated by reference. Many tumor types have been shown to exhibit this phenomenon. See, e.g., Frost et al., Cancer Res. 43: 125 (1983).

It appears that tum⁻ variants fail to form progressive tumors because they initiate an immune rejection process. The evidence in favor of this hypothesis includes the ability of "tum⁻" variants of tumors, i.e., those which do not normally form tumors, to do so in mice with immune systems suppressed by sublethal irradiation, Van Pel et al., Proc. Natl. Acad. Sci. USA 76: 5282-5285 (1979); and the observation that intraperitoneally injected tum⁻ cells of mastocytoma P815 multiply exponentially for 12-15 days, and then are eliminated in only a few days in the midst of an influx of lymphocytes and macrophages (Uyttenhove et al., J. Exp. Med. 152: 1175-1183 (1980)). Further evidence includes the observation that mice acquire an immune memory which permits them to resist subsequent challenge to the same tum⁻ variant, even when immunosuppressive amounts of radiation are administered with

the following challenge of cells (Boon et al., Proc. Natl. Acad. Sci. USA 74: 272-275 (1977); Van Pel et al., supra; Uyttenhove et al., supra).

5 Later research found that when spontaneous tumors were subjected to mutagenesis, immunogenic variants were produced which did generate a response. Indeed, these variants were able to elicit an immune protective response against the original tumor. See Van Pel et al., J. Exp. Med. 157: 1992-2001 (1983). Thus, it has been shown that it is possible to
10 elicit presentation of a so-called "tumor rejection antigen" in a tumor which is a target for a syngeneic rejection response. Similar results have been obtained when foreign genes have been transfected into spontaneous tumors. See Pearson et al., Cancer Res. 48: 2975-1980 (1988) in this
15 regard.

A class of antigens has been recognized which are presented on the surface of tumor cells and are recognized by cytotoxic T cells, leading to lysis. This class of antigens will be referred to as "tumor rejection antigens" or "TRAs" hereafter. TRAs may or may not elicit antibody responses. The extent to which these antigens have been studied, has been via cytolytic T cell characterization studies, in vitro i.e., the study of the identification of the antigen by a particular cytolytic T cell ("CTL" hereafter) subset. The subset
20 proliferates upon recognition of the presented tumor rejection antigen, and the cells presenting the antigen are lysed. Characterization studies have identified CTL clones which specifically lyse cells expressing the antigens. Examples of this work may be found in Levy et al., Adv. Cancer Res. 24: 1-59 (1977); Boon et al., J. Exp. Med. 152: 1184-1193 (1980); Brunner et al., J. Immunol. 124: 1627-1634 (1980); Maryanski et al., Eur. J. Immunol. 124: 1627-1634 (1980); Maryanski et al., Eur. J. Immunol. 12: 406-412 (1982); Palladino et al., Canc. Res. 47: 5074-5079 (1987). This type of analysis is
25 required for other types of antigens recognized by CTLs, including minor histocompatibility antigens, the male specific H-Y antigens, and the class of antigens referred to as "tum-"
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antigens, and discussed herein.

A tumor exemplary of the subject matter described supra is known as P815. See DePlaen et al., Proc. Natl. Acad. Sci. USA 85: 2274-2278 (1988); Szikora et al., EMBO J 9: 1041-1050 (1990), and Sibille et al., J. Exp. Med. 172: 35-45 (1990), the disclosures of which are incorporated by reference. The P815 tumor is a mastocytoma, induced in a DBA/2 mouse with methylcholanthrene and cultured as both an in vitro tumor and a cell line. The P815 line has generated many tum⁻ variants following mutagenesis, including variants referred to as P91A (DePlaen, supra), 35B (Szikora, supra), and P198 (Sibille, supra). In contrast to tumor rejection antigens - and this is a key distinction - the tum⁻ antigens are only present after the tumor cells are mutagenized. Tumor rejection antigens are present on cells of a given tumor without mutagenesis. Hence, with reference to the literature, a cell line can be tum⁺, such as the line referred to as "P1", and can be provoked to produce tum⁻ variants. Since the tum⁻ phenotype differs from that of the parent cell line, one expects a difference in the DNA of tum⁻ cell lines as compared to their tum⁺ parental lines, and this difference can be exploited to locate the gene of interest in tum⁻ cells. As a result, it was found that genes of tum⁻ variants such as P91A, 35B and P198 differ from their normal alleles by point mutations in the coding regions of the gene. See Szikora and Sibille, supra, and Lurquin et al., Cell 58: 293-303 (1989). This has proved not to be the case with the TRAs of this invention. These papers also demonstrated that peptides derived from the tum⁻ antigen are presented by the L^d molecule for recognition by CTLs. P91A is presented by L^d, P35 by D^d and P198 by K^d.

PCT application PCT/US92/04354, filed on May 22, 1992 assigned to the same assignee as the subject application, teaches a family of human tumor rejection antigen precursor coding genes, referred to as the MAGE family. Several of these genes are also discussed in van der Bruggen et al., Scienc 254: 1643 (1991). It is now clear that the various genes of the MAGE family are expressed in tumor cells, and can

serve as markers for the diagnosis of such tumors, as well as for their purposes discussed therein. See also Traversari et al., Immunogenetics 35: 145 (1992); van der Bruggen et al., Science 254: 1643 (1991). The mechanism by which a protein is processed and presented on a cell surface has now been fairly well documented. A cursory review of the development of the field may be found in Barinaga, "Getting Some 'Backbone': How MHC Binds Peptides", Science 257: 880 (1992); also, see Fremont et al., Science 257: 919 (1992); Matsumura et al., Science 257: 927 (1992); Latron et al., Science 257: 964 (1992). These papers generally point to a requirement that the peptide which binds to an MHC/HLA molecule be nine amino acids long (a "nonapeptide"), and to the importance of the first and ninth residues of the nonapeptide.

Studies on the MAGE family of genes have now revealed that a particular nonapeptide is in fact presented on the surface of tumor cells, and that the presentation of the nonapeptide requires that the presenting molecule be HLA-A1. Complexes of the MAGE-1 tumor rejection antigen (the "TRA" or nonapeptide") leads to lysis of the cell presenting it by cytolytic T cells ("CTLs"). This observation has both diagnostic and therapeutic implications, as discussed herein.

Research presented in, e.g., U.S. patent application Serial No. 07/938,334 filed August 31, 1992, which is the parent of the subject application showed that, when comparing homologous regions of various MAGE genes to the region of the MAGE-1 gene coding for the relevant nonapeptide, there is a great deal of homology. Indeed, these observations lead to one of the aspects of the invention disclosed and claimed therein, which is a family of nonapeptides all of which have the same N-terminal and C-terminal amino acids. These nonapeptides were described as being useful for various purposes which includes their use as immunogens, either alone or coupled to carrier peptides. Nonapeptides are of sufficient size to constitute an antigenic peptide, and the antibodies generated thereto were described as being useful for identifying the nonapeptide, either as it exists alone, or

as part of a large polypeptide.

The nonapeptides were described as being useful for identifying various HLA subtypes on the surface of tumor cells, such as melanomas. Via this ability they served both as diagnostic markers and as therapeutic agents. These features are discussed infra.

The nucleic acid sequences which code for the nonapeptides were also described therein. These nucleic acid sequences were described as also being useful as diagnostic probes for tumor presence.

The application also showed how it had been found that a cellular model could be used, wherein a non-human cell can be transfected with a nucleic acid sequence coding for a human HLA molecule. The resulting transfectant could then be used to test for nonapeptide specificity of the particular HLA molecule, or as the object of a second transfection with a MAGE gene. The co-transfectant could be used to determine whether the particular MAGE based TRA is presented by the particular HLA molecule.

The present invention deals with one of the peptides described in the earlier of the two parent application. Specifically, the nonapeptide

Glu Val Asp Pro Ile Gly His Leu Tyr

derived from the tumor rejection antigen precursor coded for by MAGE-3 has now been found to be presented by human leukocyte antigen HLA-A1. This finding and the ramifications thereof are described in the disclosure which follows.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 outlines the procedure by which a 300 base pair fragment of MAGE-1 gene was identified as coding for the relevant tumor rejection antigen.

Figure 2 shows lytic studies in which cells were incubated with various MAGE 1 peptides.

Figure 3 compares lysis of mouse cells transfected with HLA-A1 genes, in the presence of the MAGE-1 nonapeptide, and when co-transfected with the sequence coding for MAGE-1.

Figure 4 compares nonapeptides from various homologous

sections of MAGE genes and the nucleic acid sequences coding for these nonapeptid .

Figure 5 shows results from a chromium release assay using CTL clone 20/38 on various cell lines.

5 Figure 6 presents the result of assays undertaken to determine antigenic specificity of CTL clone 20/38.

Figure 7 shows the results obtained when a TNF release assay was carried out on various transfected cells.

10 Figure 8 sets forth the results of a lytic assay using the peptide of the invention.

SEQ ID NOS: 1-9 show homologous nonapeptides from MAGE genes and the nucleic acid sequences coding for these.

15 **DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS**

Example 1

The 2.4 Kb BamIII fragment, described by van der Bruggen, et al., Science 254: 1643 (1991), the disclosure of which is incorporated by reference, is known to contain only exons 2 and 3 of the gene coding for MAGE-1 protein. The fragment transfers expression of antigen MZ2-E to E⁻ antigen loss cell line variant MZ2-MEL.2.2, and leads to lysis of the transfectants by E⁺ CTLs. Previous work by DePlaen et al., Proc. Natl. Acad. Sci. USA 85: 2274 (1988), and Chomez et al., Immunogenetics 35: 241 (1990), had established that small gene fragments containing antigen peptide coding sequences regularly express those antigens, even when not transfected in the form of expression vectors. In view of these observations, experiments were carried out with smaller fragments of the 2.4 kb fragment. Various restriction enzymes were used to cut the 2.4 kb fragment into smaller fragments. The resulting, smaller fragments were cloned into plasmid vector pTZ18R. A 300 base pair fragment taken from exon 3 was obtained via polymerase chain reaction ("PCR") amplification, using oligonucleotides VDB 14:

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5'-CAGGGAGCCAGTCACAAAG-3'

and CHO 9:

5'-ACTCAGCTCCTCCAGATTT-3'.

These primers amplify a 300 base pair fragment of MAGE 1, between positions 422 and 722 of exon 1. The fragment was cloned into expression vector PSVK3. The new constructs were cotransfected with plasmid pSVtkneoB into the MZ2.MEL 2.2 cell lines. This was accomplished using the calcium phosphate precipitation method (Traversari et al., Immunogenetics 35: 145 (1992); Wölfel et al., Immunogenetics 26: 178 (1987)), using 4×10^6 cells and 3 ug of pSVtkneoB (Nicolas et al., CSH Conf. Cell Prolif 10: 469 (1983)), and 30 ug of the pTZ18R or PSVK3 constructs. The transfectants were then selected in medium containing neomycin analog G418. Fifteen days after transfection, resistant cells were tested for their ability to stimulate TNF production by the anti-E antigen CTL 82/30. This was accomplished by adding 100 ul samples, containing 1500 CTL 82/30 to 4×10^4 transfected cells. Supernatant samples (50 ul) were harvested and added to 3×10^4 WEHI 164 clone 13 cells (Espevik et al., J. Immunol. Meth. 95: 99 (1986), to evaluate TNF presence. Mortality of WEHI cells was estimated 24 hours later, using an MTT colorimetric assay as per, e.g., Traversari et al., supra.

As shown in figure 1, these experiments identified a 300 base pair fragment from MAGE-1 exon 3 capable of efficient transferring of expression of antigen MZ2E.

Example 2

The MAGE-1 gene belongs to a family of several highly related genes. See van der Bruggen et al., supra. Prior experiments had noted that MAGE-2 and MAGE-3 did not direct expression of antigen MZ2-E. As the 300 base pair fragment clearly did, the homologous sections of MAGE-2 and MAGE-3 genes were compared to the 300 base pair fragment. Differences were clear, and several 15 amino acid peptides were synthesized, using F-moc for transient N-terminal protection, in accordance with Atherton et al., J. Chem. Soc. 1: 538 (1981). The peptides were purified by C-18 reverse phase HPLC, and characterized by amino acid analysis.

Once the peptides were secured, they were tested in lysis

assays, using the chromium release methodology of Boon et al., J. Exp. Med. 152: 1184 (1980). Briefly, 1000 ^{51}Cr labeled E⁻ target cells were incubated in 96 well microplates, using various concentrations of peptides for 30 minutes at 37°C.

5 An equal volume of CTL containing sample was added (cell line 82/30), the number of CTLs being five times that of their target. Chromium release was measured after four hours. Sensitization of E⁻ cells to lysis by the anti E CTLs was observed with a peptide that corresponds to codons 158-172 of
10 the large open reading frame of MAGE-1. Shorter peptides were prepared and efficient lysis was observed with peptide: Glu Ala Asp Pro Th Gly His Ser Tyr.

 The results, shown in figure 2, demonstrate that the first and ninth amino acids were critical for binding and
15 effecting lysis. This is in accordance with prior reports stating that MHC-I molecules generally are bound by nonapeptides (Rotzschke et al., Nature 348: 252 (1990)). Figure 2 also shows that half maximum lysis was obtained at a peptide concentration of 5nM.

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Example 3

 Experiments were carried out to determine what molecule presented the relevant MAGE-1 antigen. To accomplish this, an HLA-A1 gene, as taught by Girdlestone, Nucl. Acids. Res. 18:
25 6701 (1990), was transfected into a mouse cell line, P1.HTR. This line is a highly transfectable variant of mouse mastocytoma cell line P815. The resulting transfectants, referred to as "P1.HTR.A1", were incubated in the presence of the nonapeptide discussed supra, using the same lysis assay.
30 Controls were also used.

 Figure 3 shows that the cell line was lysed, showing that a model has been developed for screening for a lytic peptide, using a non-human cell.

 In experiments not described herein, similar results were
35 obtained with COS cells.

 Additional experiments were also carried out, in which cell line P1.HTR A1 was transfected with MAGE-1 cDNA. When

the lytic assay of Example 2 was carried out with this co-transfected cells, it was found that they were also lysed.

Example 4

5 Given the homology of the various genes within the MAGE family, a comparison was carried out to identify similarities amongst the homologous regions of the genes. These regions are shown in figure 4. These peptides and the nucleic acid sequences coding for them, are not identical, but show a great deal of homology, especially the identical first and ninth residues.

Example 5

This example, and examples 6-8 which follow, correspond to examples 37-40 of copending application Serial number 08/037,230 filed on March 26, 1993.

15 A cytolytic CTL clone "20/38" was obtained from peripheral blood lymphocytes of melanoma patient MZ2. This clone is described by Van den Eynde et al., Int. J. Cancer 44: 634-640 (1989), the disclosure of which is incorporated by reference. The CTL clone was isolated following Herin et al.,
20 Int. J. Cancer 39: 390-396 (1987), which is incorporated by reference. The assay is described herein, however. Autologous melanoma cells were grown in vitro, and then resuspended at 10^7 cells/ml in DMEM, supplemented with 10% HEPES and 30% FCS, and incubated for 45 minutes at 37°C with
25 200 μ Ci/ml of $\text{Na}^{(51)\text{Cr}}\text{O}_4$. Labelled cells were washed three times with DMEM, supplemented with 10 mM HEPES. These were then resuspended in DMEM supplemented with 10 mM HEPES and 10% FCS, after which 100 μ l aliquots containing 10^3 cells, were distributed into 96 well microplates. Samples of the CTL
30 clone were added in 100 μ l of the same medium, and assays were carried out in duplicate. Plates were centrifuged for four minutes at 100g, and incubated for four hours at 37°C in a 5.5% CO_2 atmosphere.

35 Plates were centrifuged again, and 100 μ l aliquots of supernatant were collected and counted. Percentage of ^{51}Cr release was calculated as follows:

$$\% \text{ } ^{51}\text{Cr release} = \frac{(\text{ER}-\text{SR})}{(\text{MR}-\text{SR})} \times 100$$

where ER is observed, experimental ^{51}Cr release, SR is spontaneous release measured by incubating 10^3 labeled cells in 200 ul of medium alone, and MR is maximum release, obtained by adding 100 ul 0.3% Triton X-100 to target cells.

Those mononuclear blood samples which showed high CTL activity were expanded and cloned via limiting dilution, and were screened again, using the same methodology.

The same method was used to test target K562 cells. When EBV-B cells were used, the only change was the replacement of DMEM medium by Hank's medium, supplemented with 5% FCS.

These experiments led to isolation of CTL clone 20/38.

Figure 5 presents the results of these assays. Specifically, it will be seen that the CTL clone lysed autologous melanoma cell line MZ2-MEL.3.0, but did not lyse EBV-B cell lines, fibroblasts, K562 or non-autologous melanoma cell line SK-MEL-29.

Example 6

Once the CTL clone was recognized as being specific for the autologous cell line, it was tested for antigenic specificity. To do this, antigen loss variants derived from patient MZ2 were tested in the same type of chromium release assay described above. These target lines were MZ2-MEL 3.0, which is D⁺, E⁺, F⁺, A⁺, MZ2-MEL.61, which is D⁻, MZ2-MEL 2.2, which is E⁻, and MZ2-MEL.4, which is F⁻. In addition to CTL clone 20/38, clones which are known to be anti-A (CTL 28/336), anti-F (CTL 76/6), and anti-E (CTL 22/13) were tested.

These results are set forth in figure 6. It will be seen that CTL clone 20/38 lysed all the cell lines leading to chromium release except D⁻ cell line MZ2-MEL.61, thus indicating that the CTL clone is anti-D. This result was confirmed, in experiments not included herein, by experiments where TNF release by the CTL clone was observed only in the presence of melanoma lines presenting antigen D.

Example 7

Once antigen D was identified as the target molecule, studies were carried out to determine the HLA type which presented it. The experiments described in example A showed that antigen D was presented by MZ2-MEL, and this cell line's HLA specificity is known (i.e., A1, A29, B37, B44, Cw6, C.cl.10). It was also known, however, that a variant of MZ2-MEL which had lost HLA molecules A29, B44 and C.cl.10 still expressed antigen D, so these could be eliminated from consideration. Studies were not carried out on lines expressing B37, as none could be found.

In all, 13 allogeneic lines were tested, which expressed either HLA-A1 (10 of 13), or Cw6 (3 of 13). The cell lines were tested for their ability to stimulate release of TNF by CTL clone 20/38, using the method of Traversari et al., Immunogenetics 35: 145-152 (1992), the disclosure of which is incorporated by reference. This assay measures TNF release via testing toxicity of supernatants on WEHI 164-13 cells.

In the assays, cell samples (3000, 10,000 or 30,000 cells) from the allogeneic lines were cultured in the presence of 1500 cells of the CTL clone, and 25 u/ml of IL-2. Twenty-four hours later, the supernatant from the culture was tested against the WEHI cells for toxicity. The results are presented in Table 1, which follows.

Eight cell lines were found to stimulate TNF release from the CTL clone 20/38. All of these lines were HLA-A1. None of the Cw6 presenting lines did so.

The cell lines were also assayed to determine MAGE expression. All eight of the lines which stimulated TNF release expressed MAGE-3, whereas the two HLA-A1 lines which were negative did not.

Table 1

Melanoma	Number of cells	TNF pg/ml				Expression of Mage-3	Expression of HLA-A1
		Exp 1		Exp 2			
		-	+ CTL 20/38	-	+ CTL 20/38		
MZ2-MEL-61.2	50000		1		4	+++	+
MZ2-MEL-ET1	50000 1666		>120 66		>120 >120	+++	+
LY-1-MEL	30000	1	>120	1	>120	+++	+
	10000	1	>120	1	>120		
	3000	<1	114	2	>120		
MI-10221	30000	<1	>120			+++	+
	10000	<1	71				
	3000	<1	74				
LY-2-MEL	30000	1	57			+++	+
	10000	1	86				
	3000	1	91				
LY-4-MEL	30000	1	>120			+++	+
	10000	1	>120				
	3000	1	>120				
SK23-MEL	30000	1	112			++++	+
	10000	1	116				
	3000	1	105				
MI-6652-MEL	30000	1	3	2	4	-	+
	10000	1	2	2	5		
	3000	1	5.2	1	5		
LB34-MEL	30000	1	>120			++++	+
	10000	1	>120				
	3000	1	>120				
LB45-MEL	30000	1	11	1	30	-	+
	10000	1	6	1	12		
	3000	1	2	<1	7		
NA-6-MEL	30000	1	77	5	98	+++	+
	10000	1	104	5	>120		
	3000	1	110	4	>120		
MI-13443-MEL	30000	1	>120			++++	+
	10000	1	>120				
	3000	1	>120				
LB5-MEL	30000	1	8	4	9	+	-
	10000	<1	5	4	11		
	3000	<1	5	1	5		
SK64-MEL	30000	1	4	2	5	7	-
	10000	1	2	1	5		
	3000	1	1	1	4		
LB33-MEL	30000			1	3.5	+++	-
	10000			1	4		
	3000			1	3		
LB73-MEL	50000		16			-	-

1500 CTL 20/38 and 25 u/ml IL2 were mixed with the indicated number of cells of the different allogeneic melanomas. 24 h later, the amount of TNF present in the supernatant was assayed by testing its cytotoxicity for WEHI-164-13 cells.

Exempl 8

In vi w of the results set forth in example 7, experiments were carried out to determine if antigen D was in fact a tumor rejection antigen derived from MAGE-3. To do this, recipient COS-7 cells were transfected with 100ng of the gene for HLA-A1 cloned into pcDNA I/Amp, and 100 ng of one of (a) cDNA for MAGE-1 cloned into pcDNA I/Amp, (b) cDNA for MAGE-2 cloned into pcDSR α , or (c) cDNA for MAGE-3 cloned into pcDSR α . The transfecting sequences were ligated into the plasmids in accordance with manufacturer's instructions. Samples of COS-7 cells were seeded, at 15,000 cells/well into tissue culture flat bottom microwells, in Dulbeco's modified Eagles Medium ("DMEM") supplemented with 10% fetal calf serum. The cells were incubated overnight at 37°C, medium was removed and then replaced by 30 μ l/well of DMEM medium containing 10% Nu serum, 400 μ g/ml DEAE-dextran, 100 μ M chloroquine, and the plasmids described above. Following four hours of incubation at 37°C, the medium was removed, and replaced by 50 μ l of PBS containing 10% DMSO. This medium was removed after two minutes and replaced by 200 μ l of DMEM supplemented with 10% of FCS.

Following this change in medium, COS cells were incubated for 24 hours at 37°C. Medium was then discarded, and 1500 cells of CTL clones 20/38 were added, in 100 μ l of Iscove medium containing 10% pooled human serum, supplemented with 25 u/ml of IL-2. Supernatant was removed after 24 hours, and TNF content was determined in an assay on WEHI cells, as described by Traversari et al., Immunogenetics 35: 145-152 (1992), the disclosure of which is incorporated by reference. These results are shown in Figure 7.

It will be seen that the CTL clone was strongly stimulated by COS7 cells transfected with HLA-A1 and MAGE-3, but not by the cells transfected with the other mage genes. This l ads to th conclusion that antigen D is a tumor rejection antigen derived from the tumor r j ction antigen pr cursor coded by g ne MAGE-3, and that this TRA is present d by HLA-A1 molecules.

Example 9

Further experiments were carried out using peptide

Glu Val Asp Pro Ile Gly His Leu Tyr

which is derived from the gene "MAGE-3".

5 The peptide was prepared in the same manner as were the
peptides of example 2. The chromium release assay described
in that example was also used. Cell line MZ2-MEL 61.2, which
is an antigen D loss variant of MZ2.MEL43 was labelled with
10 ⁵¹Cr, and was then tested with antigen D specific cytolytic
cell clone CTL 20/38, and varying concentrations of the
peptide. MZ2-MEL61.2 and CTL 20/38 were combined in a 1.5:1
ratio, together with the peptide at varying concentrations.
The mixture was incubated for four hours, after which chromium
release was measured. As a control, chromium labelled MZ2-
15 MEL.43 was used.

The results, presented in figure 8, show that the peptide
does act as a tumor rejection antigen in that the cytolytic T
cell clones recognize and lyse the targeted cells.

20 The foregoing examples show that a nonapeptide derived
from MAGE-3 is presented by HLA-A1 molecules, and cells
presenting the complex of HLA-A1 and the nonapeptide are
recognized and lysed by specific CTL cells. This observation
indicates that the nonapeptide of the invention may be used
25 both therapeutically and diagnostically.

In the case of the latter category of use, the
nonapeptide may be used, for example, to identify tumors
expressing a particular HLA molecule, or cancer cells per se.
One contacts a cancer cell containing sample or a tumor cell
30 with the nonapeptide which binds thereto, and combines the
material with a CTL sample specific for the complex. If lysis
ensues, then the tumor/cancer cell can be identified as HLA-A1
presenters.

Therapeutically, there are two major ways in which the
35 nonapeptide may be used. In an in vivo therapeutic approach,
the nonapeptide may be administered in a way which targets it
to tumors to be treated. This can be done via direct

injection, time release administration, coupling to tumor specific antibodies, and so forth. Upon binding to HLA-A1 molecules, there is a CTL response, leading to lysis of the tumor. Of course, in such a therapeutic approach, the nonapeptide is administered in an amount sufficient to lead to lysis of the tumor. This amount will vary, based upon the particular patient, the type and size of the tumor, and so forth.

An "in vitro" form of therapy is also contemplated. As indicated supra, when HLA-A1 molecules bind to the MAGE-3 derived nonapeptide, if contacted with the CTLs specific for the peptide/HLA complex, a CTL proliferative response occurs. As the CTLs are the agents of tumor lysis in vivo, the resulting expanded populations may be administered to the patient. The CTLs can be expanded by using the patient's own blood or any other source of CTLs, or by contact to samples of peptide specific CTLs which have previously been established. In this regard, note that CTL 20/38, discussed supra had been available for some time as was the methodology for its development.

Therapies of the type described herein are particularly useful for melanoma. Analysis of samples has shown that about 26% of the caucasian population at large presents HLA-A1 allele. Thus, at the least, 26% of the caucasian melanoma population may be considered as potential subjects for therapy with the peptide. The patients may also be treated with proliferative cells which have complexes of HLA-A1 and the nonapeptide presented on their surface.

The nucleic acid sequences, as indicated, may be used in a variety of ways. MAGE genes are expressed in tumors, and thus the nucleic acid sequences may be used as probes to identify tumor cells. This can be accomplished via labelled hybridization probes, PCR, or any of the various nucleic acid probe based assays known to the art.

The development of the non-human cell lines described herein presents a unique way to carry out some of the features of the invention described herein. The examples show, e.g.,

that the CTLs recognize the complex of HLA and nonapeptide, and do not appear to differentiate between the cell types which present the complexes. Thus, the isolated, non-human cell lines of the invention can be used to generate CTLs, and to identify their presence in human samples.

As indicated, the invention also involves isolated non-human cell lines transfected with both an HLA-A1 gene, and a sequence coding for the nonapeptide. One is not limited to transfection with one HLA coding gene and one MAGE peptide, and indeed the invention contemplates polytransfected cells, which may contain more than one HLA gene and more than one MAGE antigen coding sequence. Given the finding that both a MAGE-1 derived nonapeptide and a MAGE-3 derived nonapeptide are presented by a common HLA molecule supports this contention. Such cells may be regarded as universal effector cells, as the presence of appropriate pairs of HLA and peptide on the surface will lead either to identification of specific CTLs of choice, or to generation of CTL proliferation in a therapeutic context. Such cells, be they cotransfected or polytransfected, may serve as vaccines when combined with a suitable adjuvant, such as those well known to the art. Treatment of various cancerous conditions, such as melanoma and breast cancer, may be carried out using these transfectants.

The terms and expressions which have been employed are used as terms of description and not of limitation, and there is no intention in the use of such terms and expressions of excluding any equivalents of the features shown and described or portions thereof, it being recognized that various modifications are possible within the scope of the invention.

(1) GENERAL INFORMATION:

(i) APPLICANTS: LUDWIG INSTITUTE FOR CANCER RESEARCH

(ii) TITLE OF INVENTION: ISOLATED NONAPEPTIDE DERIVED
FROM MAGE-3 GENE AND PRESENTED BY HLA-A1,
AND USES THEREOF

(iii) NUMBER OF SEQUENCES: 9

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: Felfe & Lynch

(B) STREET: 805 Third Avenue

(C) CITY: New York City

(D) STATE: New York

(E) COUNTRY: USA

(F) ZIP: 10022

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Diskette, 5.25 inch, 360 kb
storage

(B) COMPUTER: IBM PS/2

(C) OPERATING SYSTEM: PC-DOS

(D) SOFTWARE: Wordperfect

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: 08/073,103

(B) FILING DATE: 07 June 1993

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: 07/938,334

(B) FILING DATE: 31 August 1992

(A) APPLICATION NUMBER: 08/037,230

(B) FILING DATE: 26 March 1993

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(C) REFERENCE/DOCKET NUMBER: LUD 293.1

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(B) TELEFAX: (212) 838-3884

19

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

(ix) FEATURE:

(A) NAME/KEY: MAGE-1 nonapeptide coding sequence

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

GAA GCA GAC CCC ACC GGC CAC TCC TAT

27

Glu Ala Asp Pro Thr Gly His Ser Tyr

5

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

(ix) FEATURE:

(A) NAME/KEY: MAGE-2 nonapeptide coding sequence

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

GAA GTG GTC CCC ATC AGC CAC TTG TAC

27

Glu Val Val Pro Ile Ser His Leu Tyr

5

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

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20

(ix) FEATURE:

(A) NAME/KEY: MAGE-21 nonapeptide coding sequence

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

5 GAA GTG GTC CGC ATC GGC CAC TTG TAG 27
Glu Val Val Arg Ile Gly His Leu Tyr
5

(2) INFORMATION FOR SEQ ID NO: 4:

10 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: genomic DNA

(ix) FEATURE:

(A) NAME/KEY: MAGE-3 nonapeptide coding sequence

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

20 GAA GTG GAC CCC ATC GGC CAC TTG TAC 27
Glu Val Asp Pro Ile Gly His Leu Tyr
5

(2) INFORMATION FOR SEQ ID NO: 5:

25 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

30 (ii) MOLECULE TYPE: genomic DNA

(ix) FEATURE:

(A) NAME/KEY: MAGE-4 nonapeptide coding sequence

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

35

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21

GAA GTG GAC CCC GCC AGC AAC ACC TAC
Glu Val Asp Pro Ala Ser Asn Thr Tyr

27

5

5 (2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

10 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

(ix) FEATURE:

(A) NAME/KEY: MAGE-41 nonapeptide coding sequence

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

15

GAA GTG GAC CCC ACC AGC AAC ACC TAC
Glu Val Asp Pro Thr Ser Asn Thr Tyr

27

5

20 (2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

25 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

(ix) FEATURE:

(A) NAME/KEY: MAGE-5 nonapeptide coding sequence

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

30

GAA GCG GAC CCC ACC AGC AAC AAC TAC
Glu Ala Asp Pro Thr Ser Asn Thr Tyr

27

5

35 (2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

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22

(B) TYPE: nucleic acid

(C) STRANDEDNESS: singl

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

5

(ix) FEATURE:

(A) NAME/KEY: MAGE-51 nonapeptide coding sequence

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

GAA GCG GAC CCC ACC AGC AAC ACC TAC

27

10

Glu Ala Asp Pro Thr Ser Asn Thr Tyr

5

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

15

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

20

(ix) FEATURE:

(A) NAME/KEY: MAGE-6 nonapeptide coding sequence

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

GAA GTG GAC CCC ATC GGC CAC GTG TAC

27

25

Glu Val Asp Pro Ile Gly His Val Tyr

5

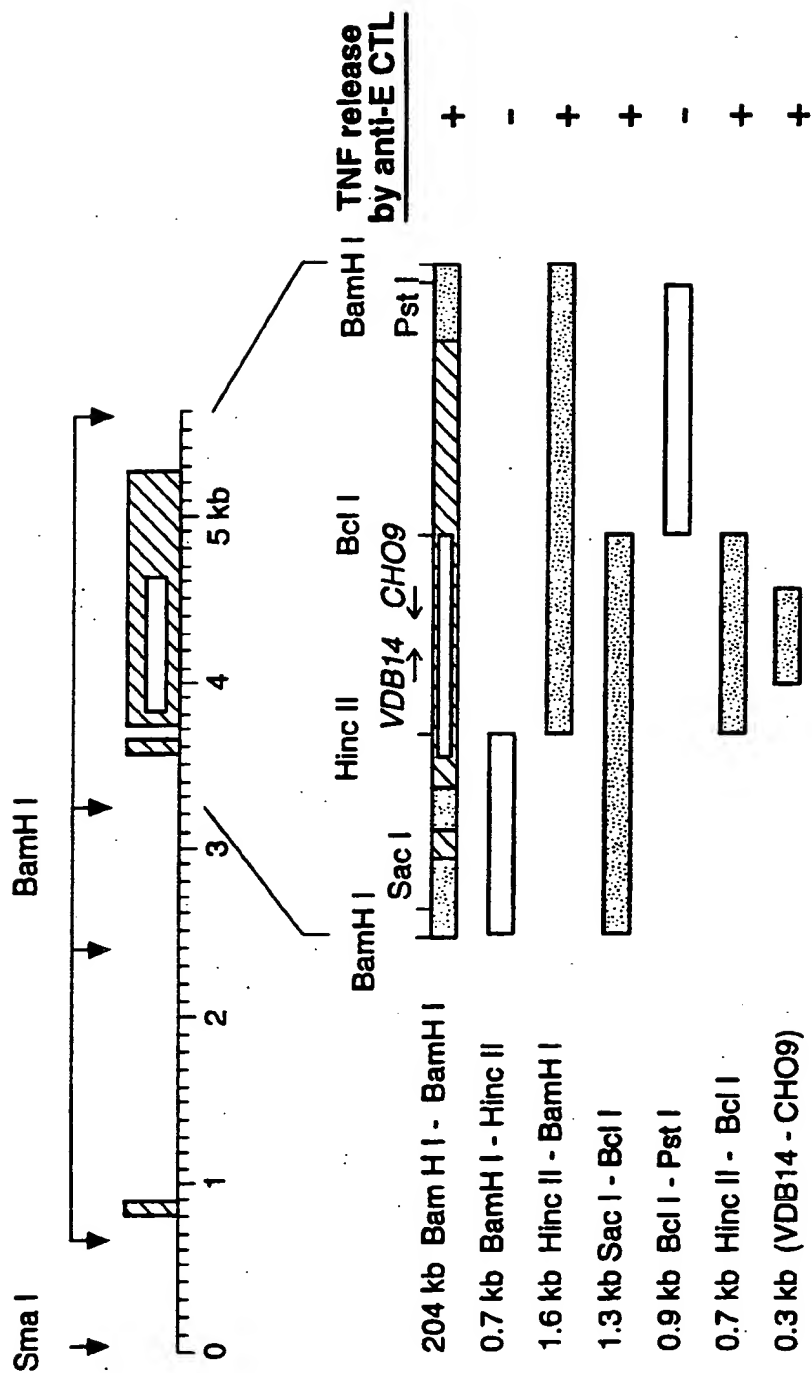
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We claim:

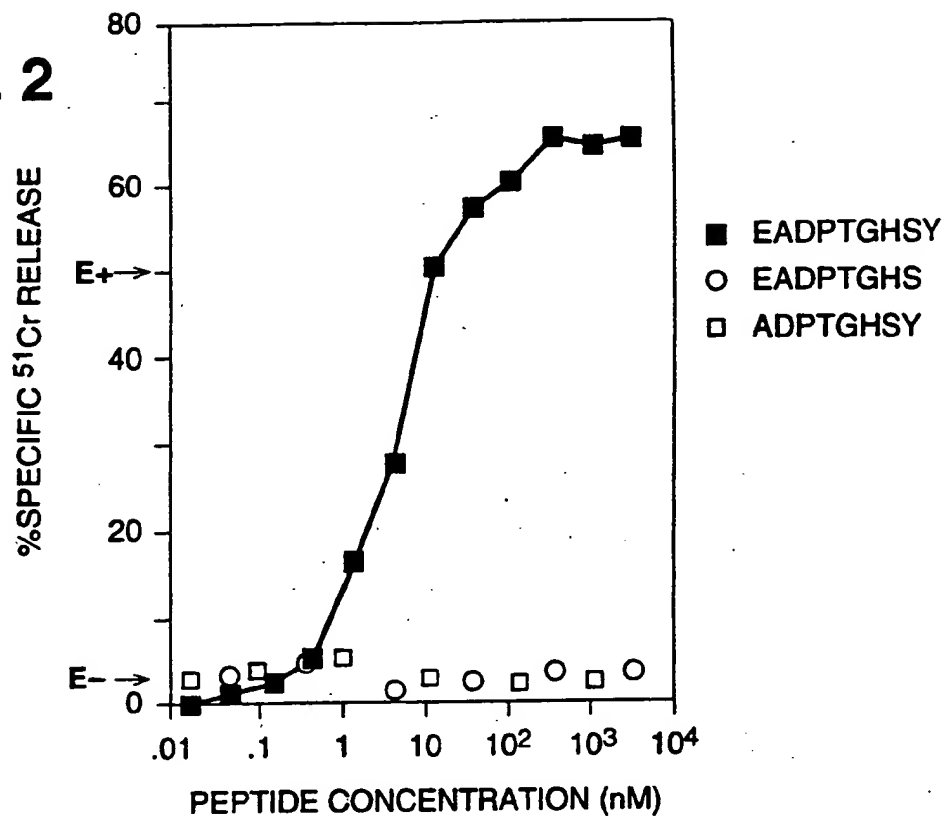
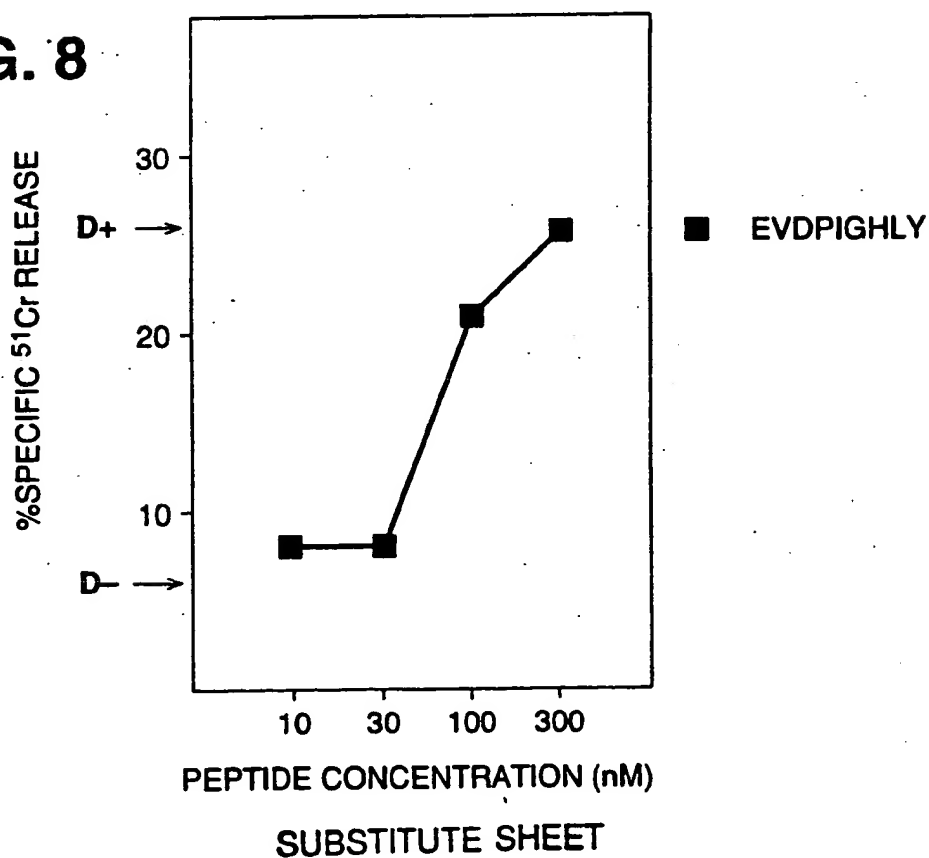
1. Isolated nonapeptide having amino acid sequence
Glu Val Asp Pro Ile Gly His Leu Tyr.
- 5 2. Isolated nucleic acid molecule coding for the nonapeptide of claim 1.
3. Method for treating a patient with a cancerous condition, wherein said cancerous condition is characterized by HLA-A1 subtype, comprising administering to said subject an
10 amount of the isolated nonapeptide of claim 1 sufficient to provoke an effective cytolytic T cell response against cancerous cells with said HLA-A1 subtype.
4. Isolated, transfected non-human cell line transfected with the nucleic acid molecule of claim 2.
- 15 5. The cell line of claim 4, wherein said cell line is a mouse cell line.
6. The cell line of claim 4, wherein said cell line is a COS cell line.
7. Method for determining a cancerous condition comprising
20 contacting a sample from a subject suspected of having a cancerous condition with a substance which specifically binds to
 - (a) nonapeptide Gly Val Asp Pro Ile Gly His Leu Tyr,
or
 - 25 (b) complexes of an HLA-A1 molecule and said nonapeptide, and
determining binding as an indication of said cancerous condition.
8. Vaccine useful in treating a cancerous condition
30 comprising the isolated nonapeptide of claim 1 and a pharmaceutically acceptable adjuvant.
9. Isolated antibody which specifically binds to the nonapeptide of claim 1.
10. The antibody of claim 9, wherein said antibody is a
35 monoclonal antibody.

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FIG. 1



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FIG. 2**FIG. 8**

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FIG. 3A

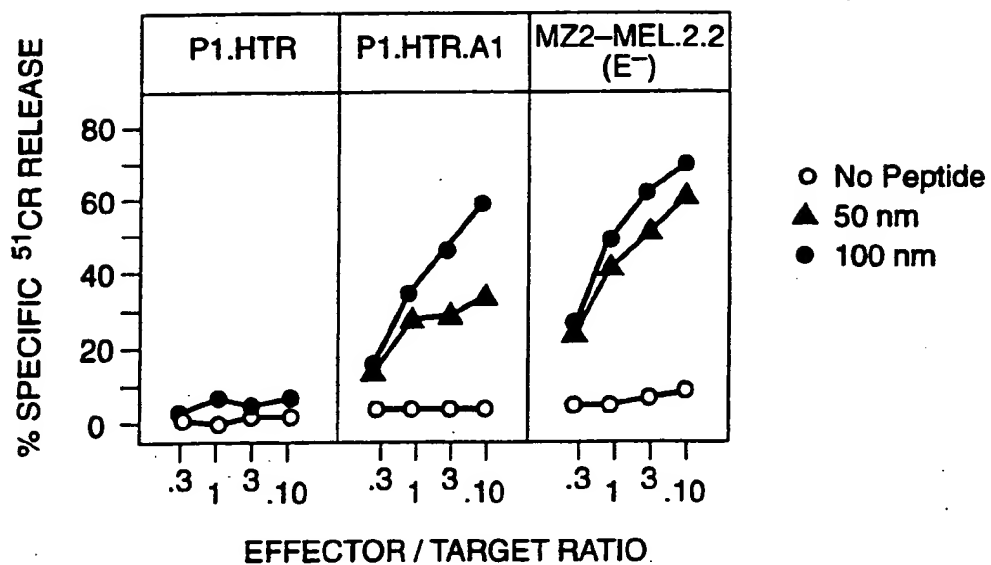
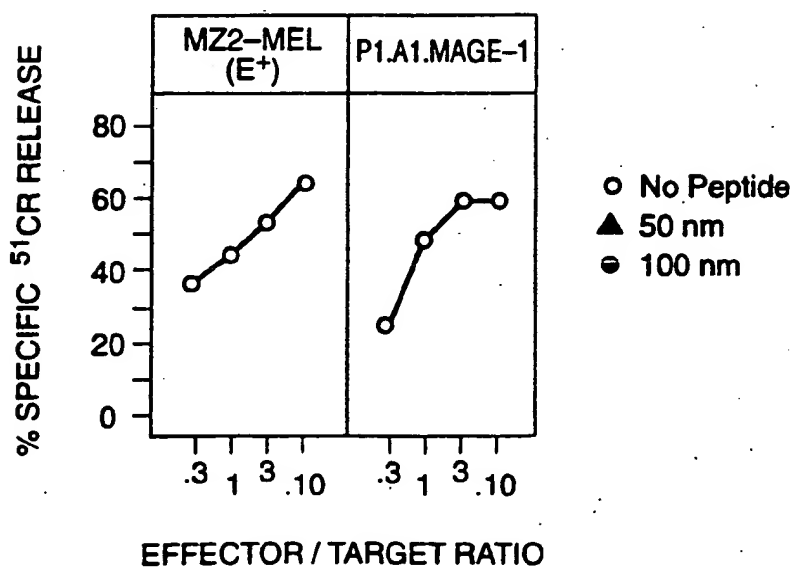


FIG. 3B



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FIG. 4

MAGE 1	E	A	D	P	T	G	H	S	Y
MAGE 2	E	V	V	P	I	S	H	L	Y
MAGE 21	E	V	V	R	I	G	H	L	Y
MAGE 3	E	V	D	P	I	G	H	L	Y
MAGE 4	E	V	D	P	A	S	N	T	Y
MAGE 41	E	V	D	P	T	S	N	T	Y
MAGE 5	E	A	D	P	T	S	N	T	Y
MAGE 51	E	A	D	P	T	S	N	T	Y
MAGE 6	E	V	D	P	I	G	H	V	Y

MAGE 1	GAA	GCA	GAC	CCC	ACC	GGC	CAC	TCC	TAT
MAGE 2	GAA	GTG	GTC	CCC	ATC	AGC	CAC	TTG	TAC
MAGE 21	GAA	GTG	GTC	CGC	ATC	GGC	CAC	TTG	TAC
MAGE 3	GAA	GTG	GAC	CCC	ATC	GGC	CAC	TTG	TAC
MAGE 4	GAA	GTG	GAC	CCC	GCC	AGC	AAC	ACC	TAC
MAGE 41	GAA	GTG	GAC	CCC	ACC	AGC	AAC	ACC	TAC
MAGE 5	GAA	GCG	GAC	CCC	ACC	AGC	AAC	ACC	TAC
MAGE 51	GAA	GCG	GAC	CCC	ACC	AGC	AAC	ACC	TAC
MAGE 6	GAA	GTG	GAC	CCC	ATC	GGC	CAC	GTG	TAC

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FIG. 5A

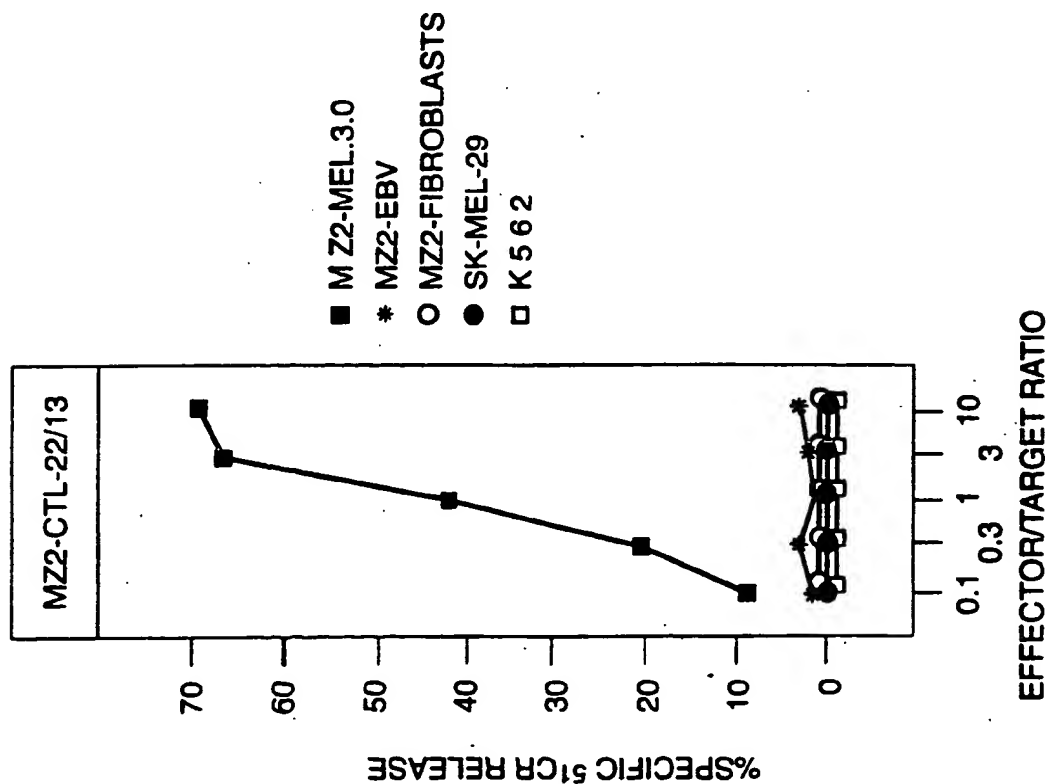
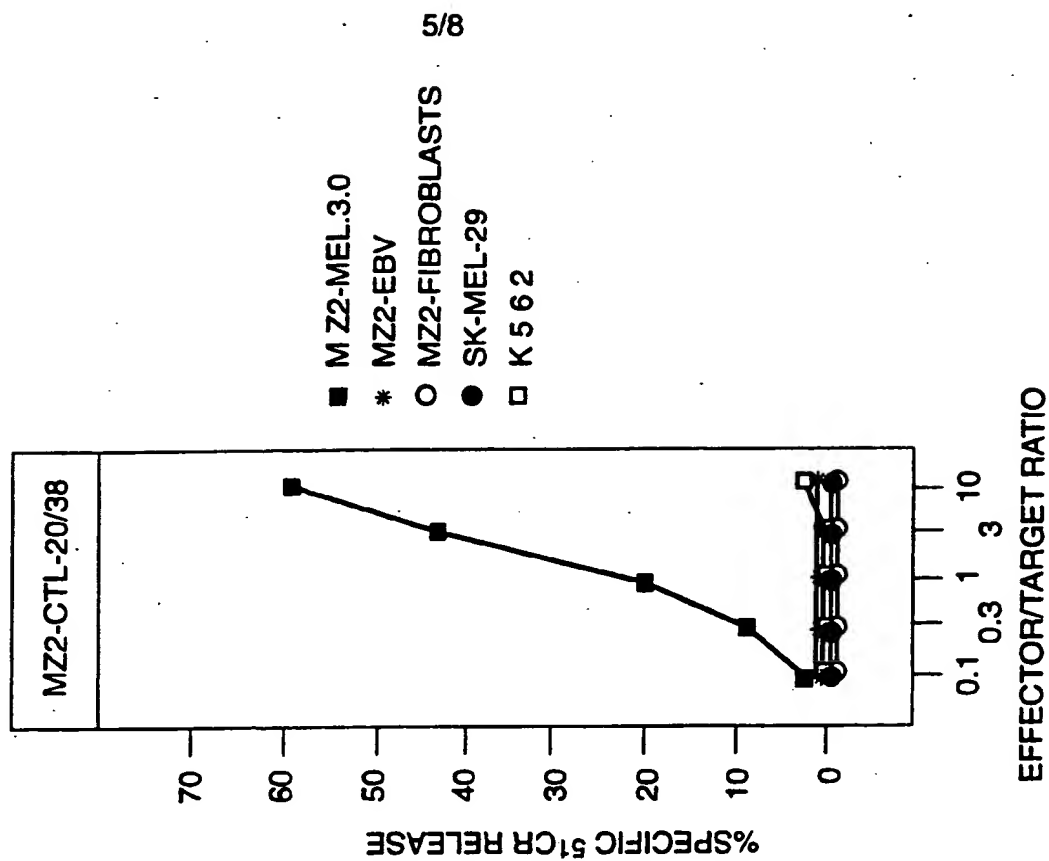
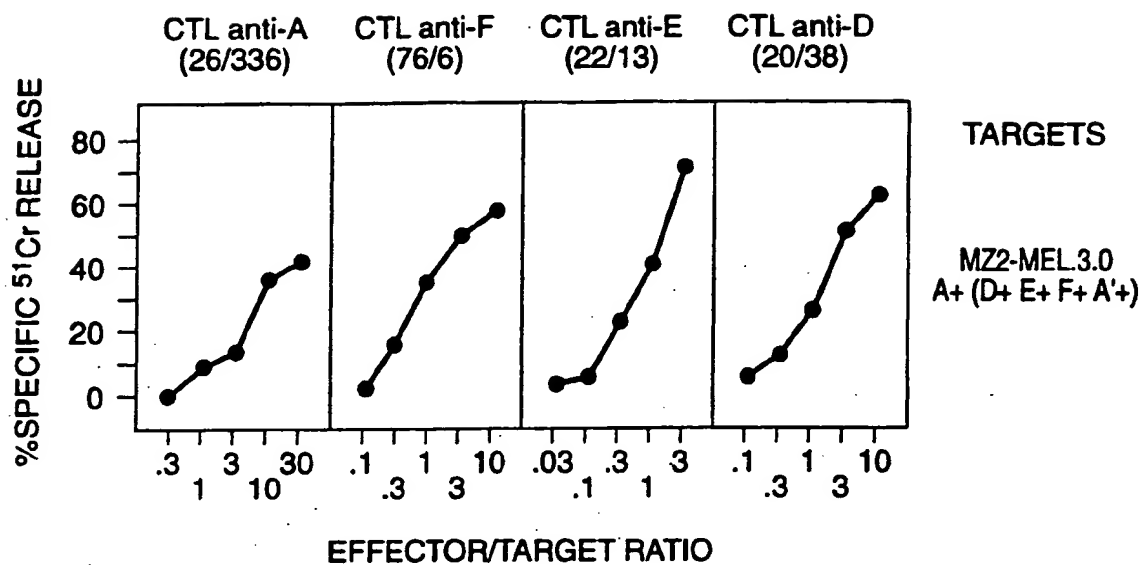
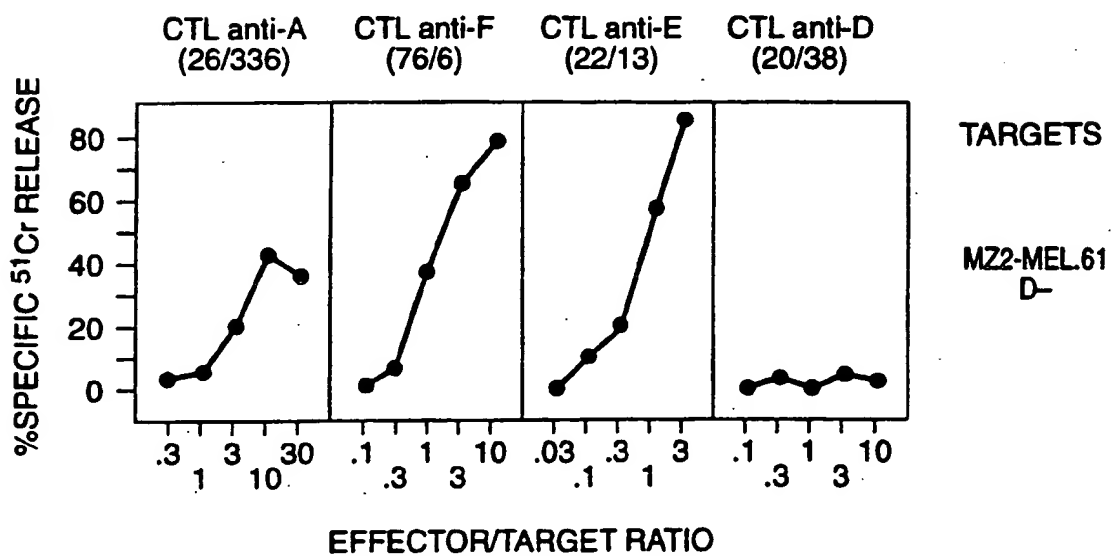


FIG. 5B

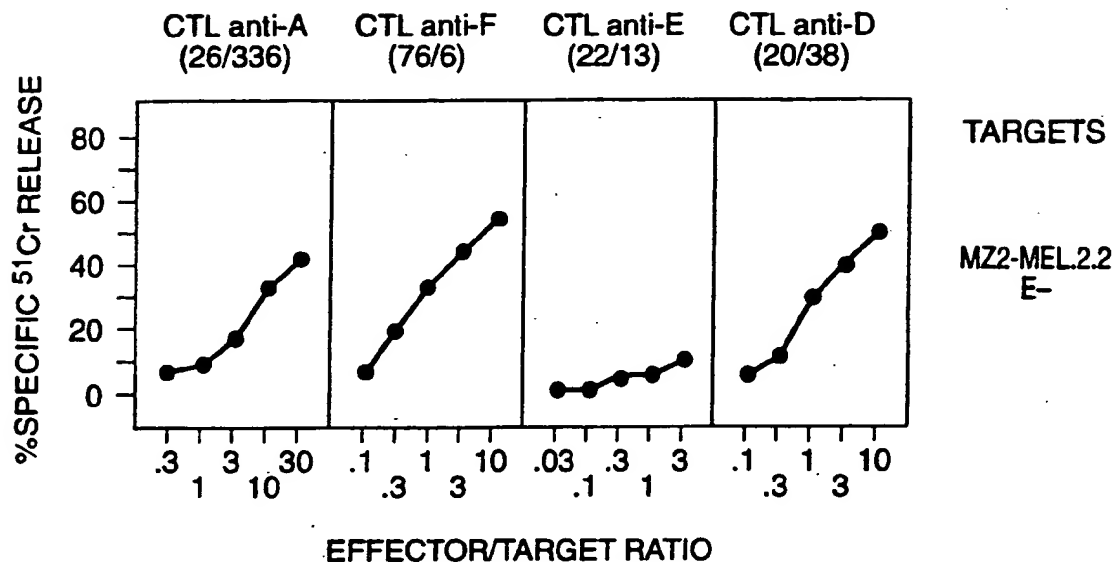
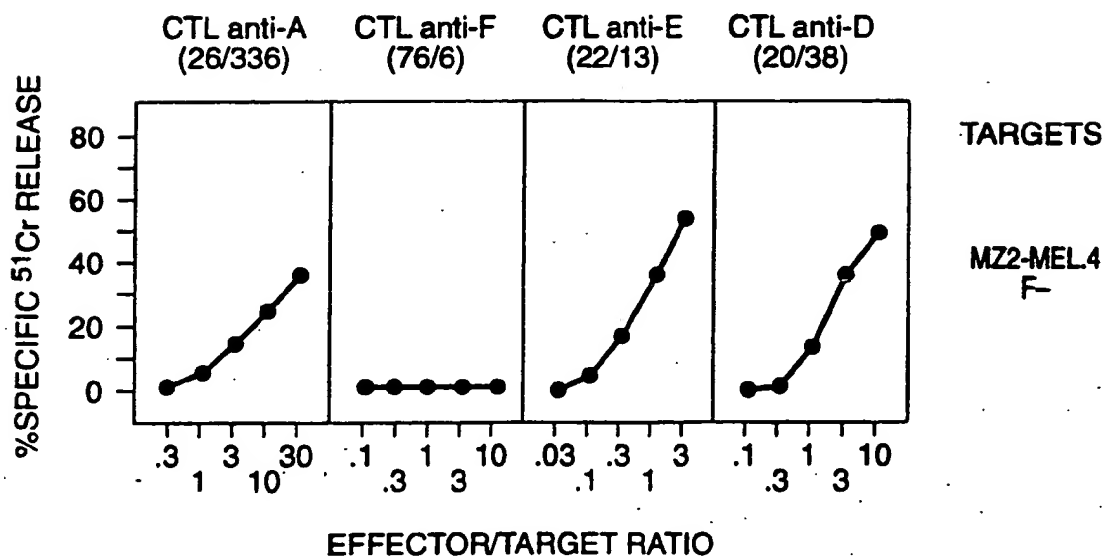


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FIG. 6A**FIG. 6B**

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FIG. 6C**FIG. 6D**

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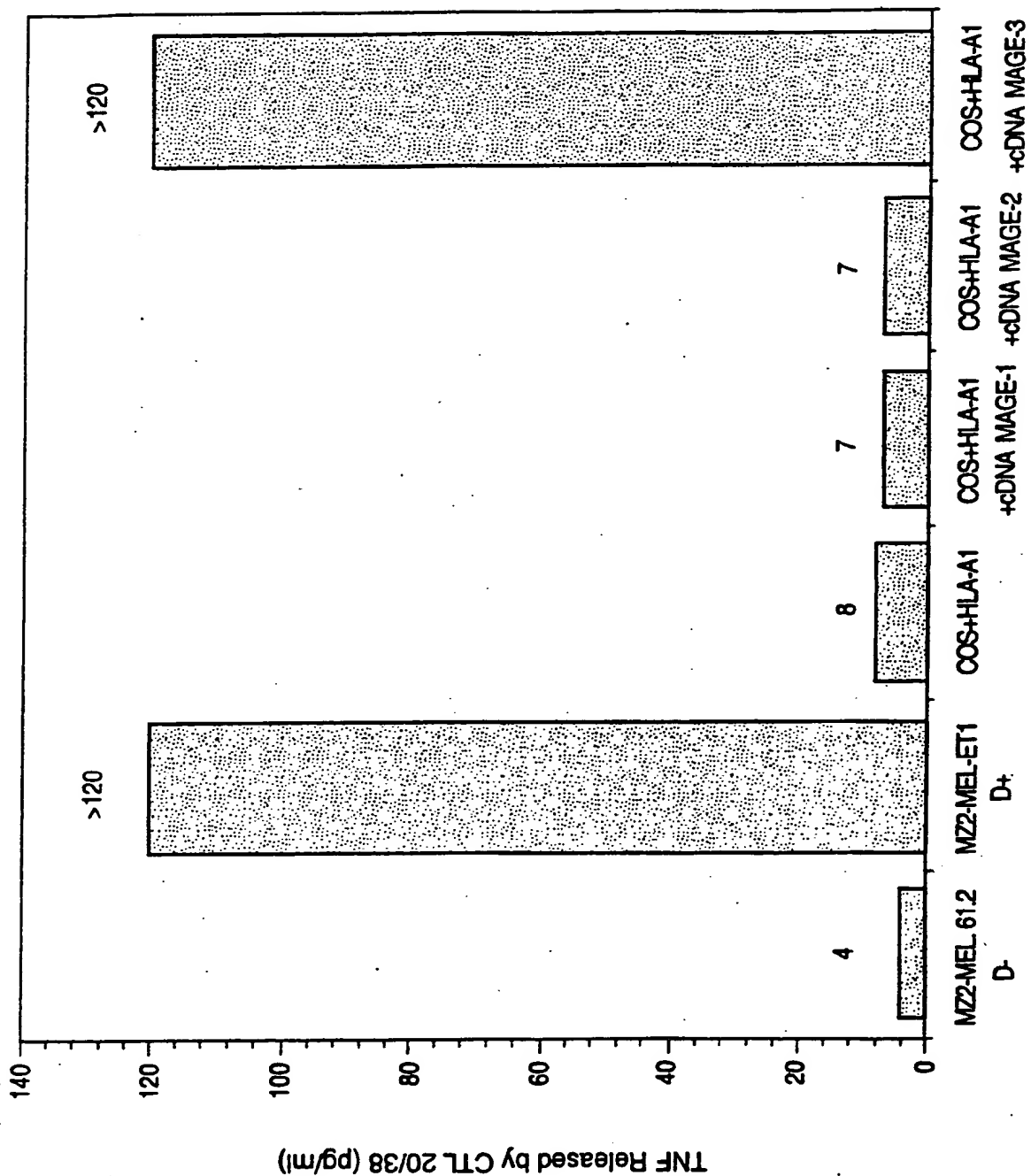


FIG. 7

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INTERNATIONAL SEARCH REPORT

International Application No.
PCT/US93/08157**A. CLASSIFICATION OF SUBJECT MATTER**

IPC(S) : A61K 35/16; C07K 15/28; 3/00, 13/00, 15/00, 17/00

US CL : 530/328, 387.1, 387.7, 388.8

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 530/328, 387.1, 387.7, 388.8

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

CAS registry, Chem abstracts, PIR, Swiss-Prot

search terms: inventors, sequence of claim 1, HLA-A1, receptor

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Science, Vol 254, issued December 1991, Bruggen et al. "A Gene Encoding an Antigen Recognized by Cytolytic T Lymphocytes on a Human Melanoma," pages 1643-1647, see entire reference.	1-10
Y,P	TibTech, Vol 11, issued February 1993, Harris et al. "Therapeutic antibodies - the coming of age," pages 42-46, see specifically page 42, column 2, lines 4-7 and column 3, lines 17-20.	1-10

☐ Further documents are listed in the continuation of Box C.
 ☐ See patent family annex.

* Special categories of cited documents:	* T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
* A* document defining the general state of the art which is not considered to be part of particular relevance	* X	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
* E* earlier document published on or after the international filing date	* Y	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
* L* document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	* Z	document member of the same patent family
* O* document referring to an oral disclosure, use, exhibition or other means		
* P* document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

22 November 1993

Date of mailing of the international search report

15 DEC 1993

Name and mailing address of the ISA/US
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Box PCT
Washington, D.C. 20231

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